

# **Incorporating relatedness in the study of molecular phenotypes for genomic epidemiology**

**Chris Holmes, Ingileif B. Hallgrímsdóttir, George Nicholson**

**Oxford Centre for Gene Function,**

**Department of Statistics,**

**University of Oxford**

## Overview

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- MoIPAGE study in genomic epidemiology
- Use of measures of **relatedness** on individuals for estimating genetic and technical variability in molecular phenotypes
- Bayesian Variance Components Models
  - illustrated for Spectral data: ClinProt MALDI-TOF data
- **Co-variance components models**

## MoIPAGE

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- MoIPAGE stands for **M**olecular **P**henotyping to **A**ccelerate **G**enomic **E**pidemiology. Funded under EU FP6.
- Take a common biological sample (fat, urine and plasma) from a set of **relateds** (twins) and molecular phenotype them on a number of platforms
  - (i) Epigenomics (genome-wide methylation profiles)
  - (ii) Gene expression (Affy)
  - (iii) Proteomics (ClinProt; peptidomics; antibody arrays)
  - (iv) Metabonomics (NMR, LCMS)
- The first phase is on quantifying the genetic (heritable) components of variation of the molecular traits *and experimental variation* (robustness) inherent in the measurement of the molecular phenotypes.

- Second stage is in integrative genomics

## Biomarkers

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- The motivation for MoIPAGE and a major research pursuit in genomic epidemiology is the search for molecular biomarkers of human disease
- Biomarker:
  - “A measurable biological trait (molecular or physiological) which associates with the onset or progression of disease”*
- Traditional biomarkers include,
  - Cholesterol, blood pressure, BMI
  - ER status (breast cancer)

## Uses of Biomarkers

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There are three major uses for biomarkers

- **Profiling** patients with increased disease risk

*“It is much more important to know the kind of patient that has a disease than to know the kind of disease a patient has” -*

$\pi(x|y)$  vrs.  $\pi(y|x)$

- Cholesterol (heart disease)
- **Prognosis** – more accurate prediction of disease progression
  - Number lymph nodes positive (breast cancer)
- **Subtyping** – towards “personalised medicine”
  - Oestrogen receptor (ER) status (breast cancer)

## Features of a Good Biomarker

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A number of features affect the utility of a biomarker (over and above prediction accuracy)

- Stability
  - both in variation of the biomarker trait over time and, as important,
  - sample storage
- Generality - coverage
- Ease of measurement: stability and accuracy of the measurement platform
- Non-invasive
- Cheap (relatively)

## Genomics and Biomarkers

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- Genomic technologies have opened up the prospect for finding new molecular markers for familial and non-familial genetic disease

## MoIPAGE Study design

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- At the first stage we are performing a **twin study** to analyse biological and technical variation
- Twins provide a powerful design for inferring genetic effects
  - blocked for in utero, dietary and socio-economic effects due to upbringing
  - known amount of genetic sharing between identical (MZ) and non-identical (DZ) twins

## Twin Study

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- Twins were contacted from St. Thomas' UK Adult Twin Registry of 10,000 twins
- The initial study has 77 twin pairs
  - 56 MZ (identical) twin pairs (31 twin pairs gave samples twice to capture longitudinal effects)
  - 21 DZ (fraternal) twin pairs
  - Fat, Urine and Plasma samples are taken
  - In total **215 samples** from the 154 individuals (**split into two aliquots, 430 aliquots**)

- The same biological samples are shipped to each technological partner for molecular phenotyping (to allow for direct comparison and integrative genomics), at least 3 technical replicates per aliquot.
- We will denote a generic molecular phenotype measurement as

$$Y_{ijkl}$$

for twin pair  $i \in \{1, \dots, 77\}$ , twin  $j \in \{1, 2\}$ , visit  $k \in \{1, 2\}$ , aliquot  $l \in \{1, 2\}$ .

## Statistical Model

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- We analyse many different molecular phenotypes
- Useful to have a common statistical structure for the model

## Twin Model

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$$Y_{ijkl} = \mu + a_{ij} + d_{ij} + c_i + e_{ij} + v_{ijk} + l_{ijkl} + b_{B(i,j,k,l)} + \epsilon_{ijkl}$$

$\mu$	:	overall mean
$a_{ij}$	:	additive genetic effect
$d_{ij}$	:	dominant genetic effect
$c_i$	:	common environmental effect
$e_{ij}$	:	individual environmental effect
$v_{ijk}$	:	individual visit effect
$l_{ijkl}$	:	aliquot effect
$b_{B(i,j,k,l)}$	:	batch effect
$\epsilon_{ijkl}$	:	residual error

## Covariance of genetic components

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- Measures of expected **relatedness** allow us to estimate the genetic (heritable) components of variation
- Since MZ twins are genetically identical  $a_{i1} = a_{i2}$  and  $d_{i1} = d_{i2}$  if twin pair  $i$  is MZ.
- DZ twins share on average half of their genetic material and

$$\text{Corr}(a_{i1}, a_{i2}) = \begin{pmatrix} 1 & 1/2 \\ 1/2 & 1 \end{pmatrix}$$

$$\text{Corr}(d_{i1}, d_{i2}) = \begin{pmatrix} 1 & 1/4 \\ 1/4 & 1 \end{pmatrix}$$

## Twin Model

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- The goal of our analysis is to partition the variability in the phenotype value into that attributable to different sources,
  - genetic ( $a_{ij}$  and  $d_{ij}$ )
  - environmental ( $c_i, e_{ij}, v_{ijk}$ )
  - technical/experimental ( $l_{ijkl}, b_{B(i,j,l,k)}$ )

- The genetic components,  $a_{ij}$  and  $d_{ij}$  and the common environment  $c_i$  are not identifiable in the likelihood and so we typically are interested in the proportion of variance attributable to

$$\text{familiality} = [a_{ij} + d_{ij} + c_i]$$

## Bayesian Model

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- The effects are unique to an individual therefore we seek to model them using hierarchical structure, for example,

$$\{a_{i1}, a_{i2}\} \sim MVN(0, \sigma_a^2 \Sigma)$$

- where  $\Sigma$  is the correlation structure and we adopt a prior

$$\sigma_a^2 \sim \pi(\cdot)$$

- and interest is on the posterior distribution  $\pi(\sigma_a^2 | Y)$  which can be obtained using MCMC (with analytic integration of the actual effects,  $a_{ij}$  etc)

## Variance Decomposition

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The total phenotypic variance is

$$\sigma_Y^2 = \sigma_a^2 + \sigma_d^2 + \sigma_c^2 + \sigma_e^2 + \sigma_v^2 + \sigma_l^2 + \sigma_b^2 + \sigma_\epsilon^2$$

and the familiarity is

$$f^2 = \frac{\sigma_a^2 + \sigma_d^2 + \sigma_c^2}{\sigma_Y^2}$$

## Gibbs Sampling

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- The joint distribution of the variance components is not known explicitly and we cannot sample from it directly.
- However, we can sample from the conditional distributions of each component (conditioned on all the others).
- The joint distribution is stationary w.r.t. the transition rule determined by the conditional distributions. Sequential draws from the conditional distributions are thus a sample from a Markov chain whose stationary distribution is the joint.

## Choice of prior distributions

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- We consider the following priors:
  - Gamma distribution on the precision,  $1/\sigma_{\star}^2 \sim \text{Gamma}(\epsilon, \epsilon)$ .
  - Uniform distribution on the standard deviation,  $\sigma_{\star} \sim \text{U}(0, C)$ .
  - Half-Cauchy distribution on the standard dev.,  $\sigma_{\star} \sim \text{hC}(s)$ .
  
- When the number of random effects that share a variance component is large (e.g. there are 154  $e_{ij} \sim N(0, \sigma_e^2)$ ) the choice of prior does not affect much the posterior distribution.
  
- For all variance components except  $\sigma_b^2$  we choose a uniform prior, but since there are only 5 batches more care needs to be taken in choosing the prior.

## Identifiability

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- The parameters  $a$ ,  $d$  and  $c$  are not all identifiable in the likelihood.
- One of the benefits of working in a Bayesian framework is that we define the model to match the underlying structure, regardless of identifiability.
- The joint posterior distributions provide us with important insight into how variance can be “transferred” between the variance components  $a$ ,  $d$  and  $c$ , giving us information about equally valid parameter values

## Identifiability

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- From a Bayesian perspective:

What you do or do not observe should not influence the model you adopt for the underlying process

- That is, you write down the model you believe underlies the data generating process and then condition on the data to hand

## Case study using ClinProt proteomics

## ClinProt MALDI-TOF Data

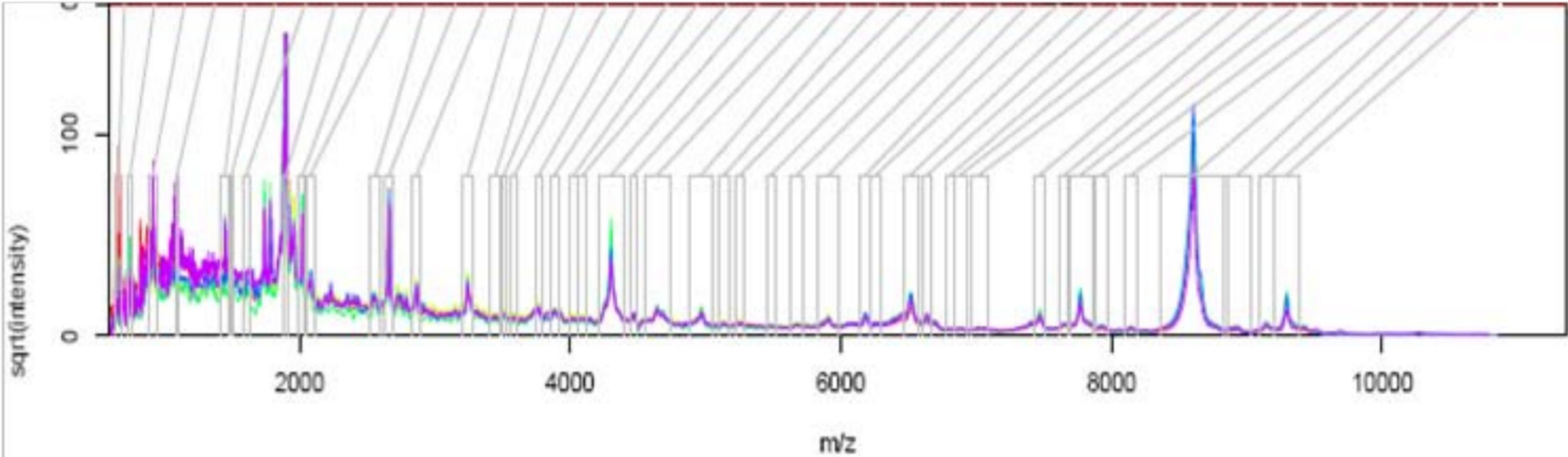
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- Magnetic beads with functional surfaces are used to bind proteins and peptides from a sample (plasma, serum or urine).
- After elution the captured proteins and peptides are analysed in a MALDI-TOF mass spectrometer.
- We perform pre-processing of the spectra, including denoising, baseline subtraction, normalisation, alignment and peak extraction.

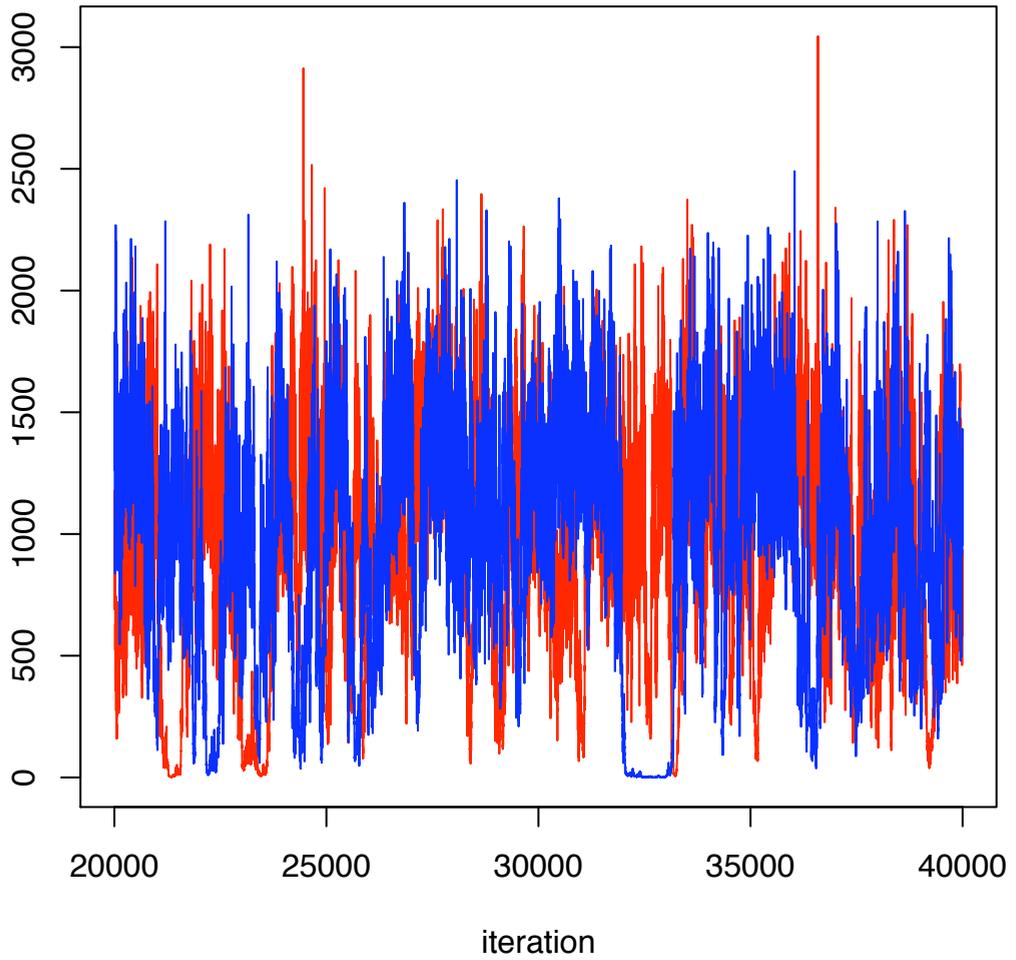
## Analysis of Peaks

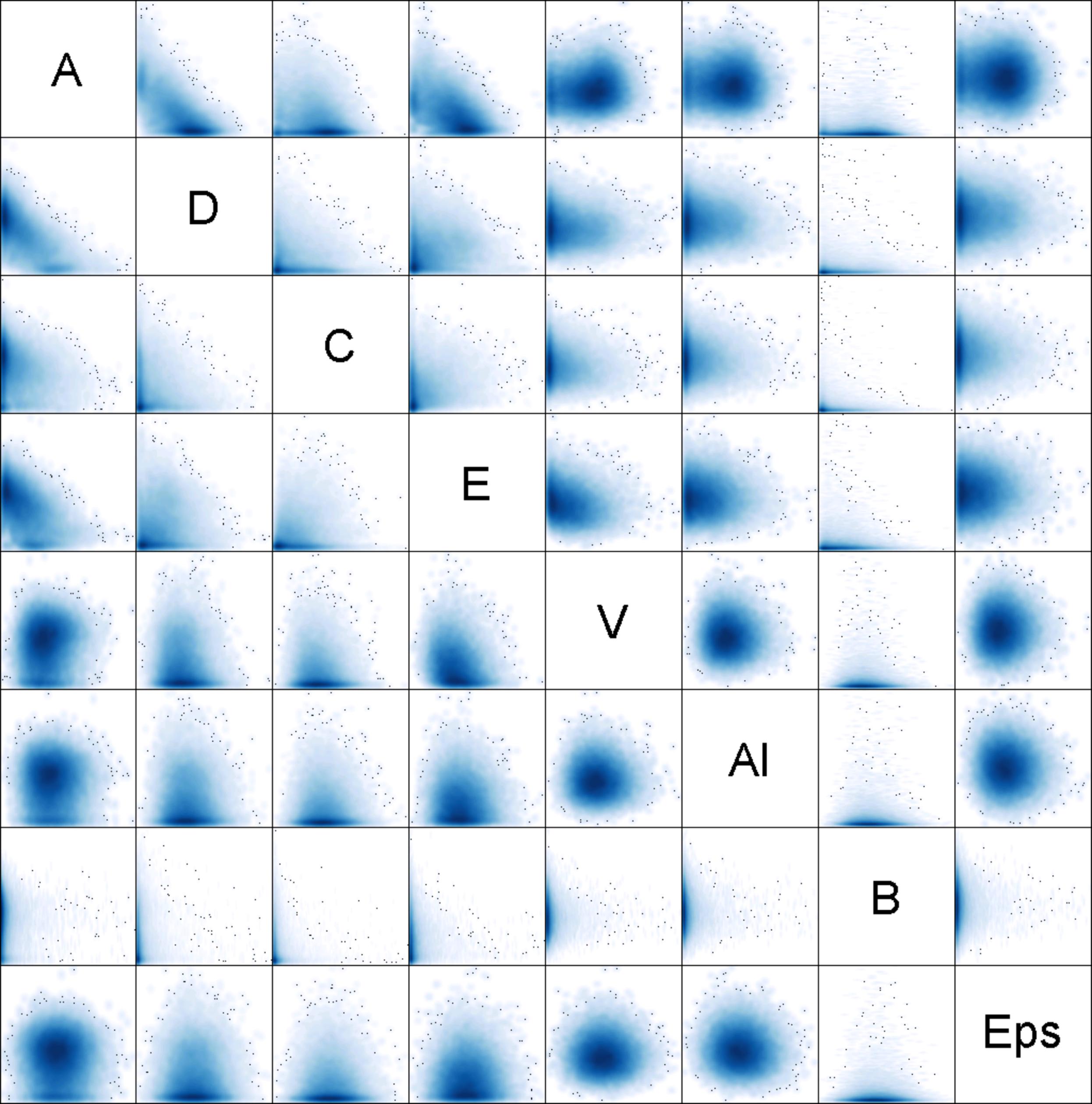
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- Peak areas are extracted and **area under the peak** is treated as phenotype  $Y_{ijkl}$
- Initially we treat each peak independently
- This is equivalent to stating that (initially) we allow for interactions between the peptides and the genetic effects
- We run our MCMC simulations and report posterior distributions on the variance components
- For example, the output for peptide abundance under one peak would look like

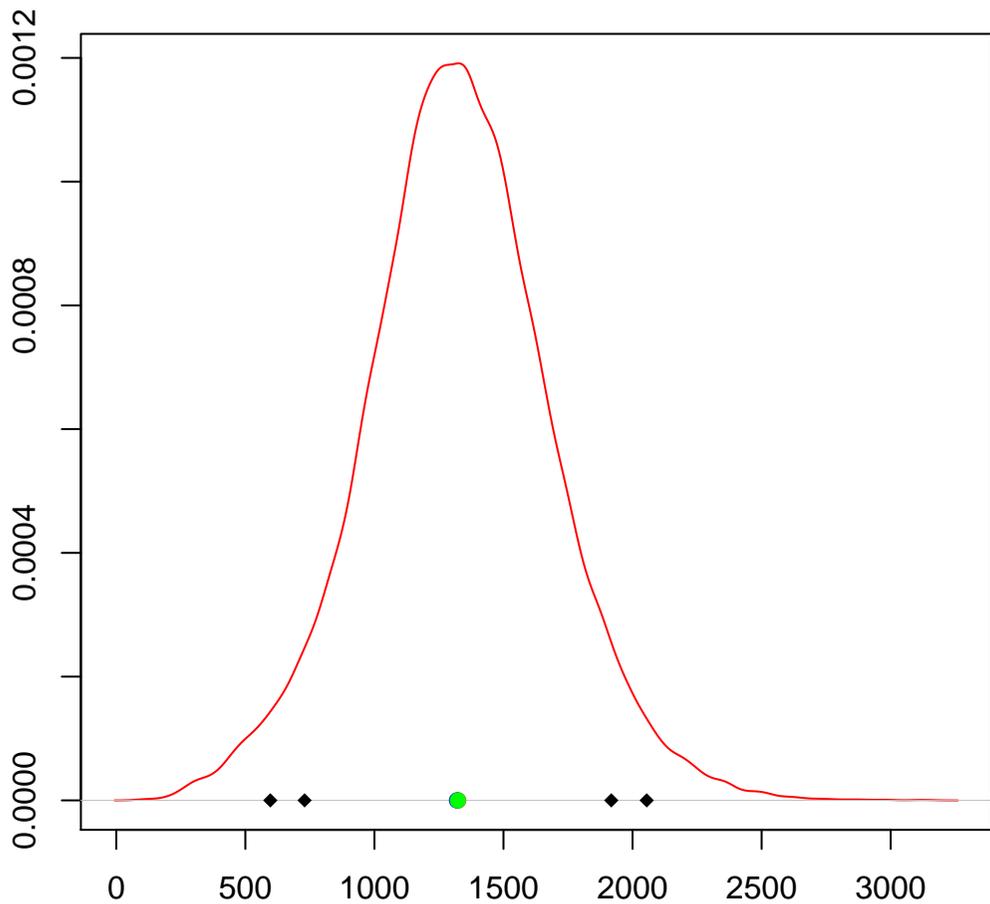


# 'VarA'

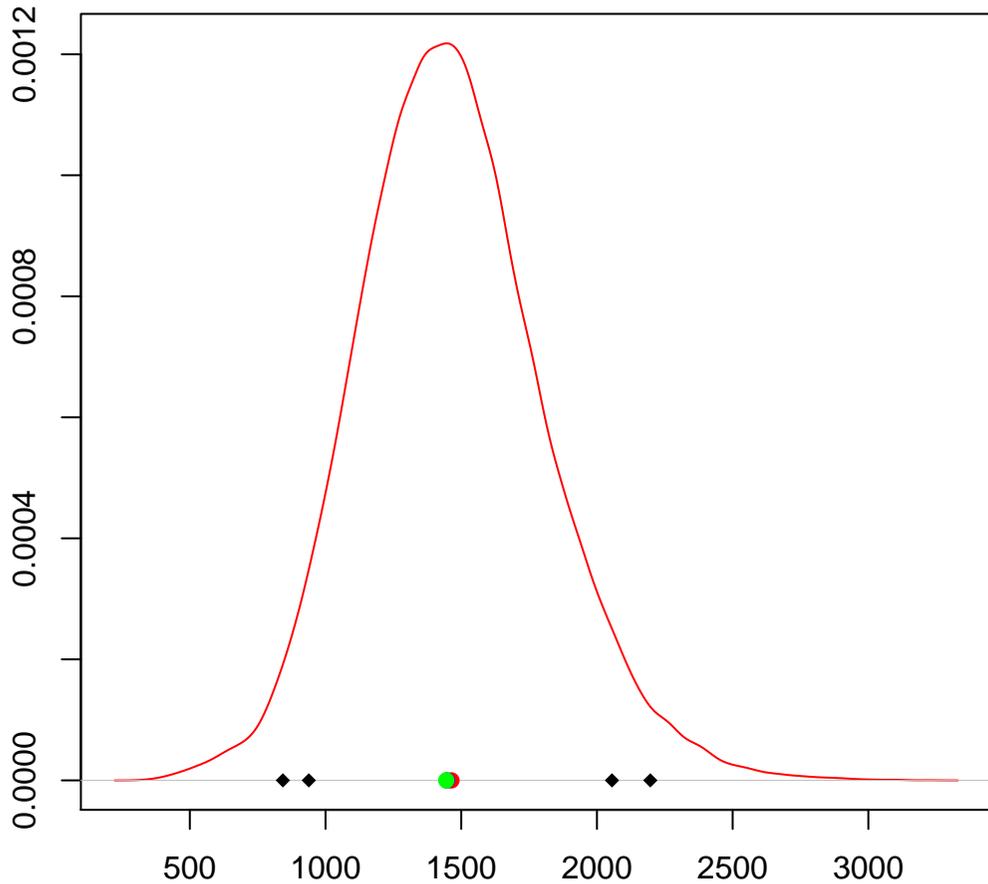




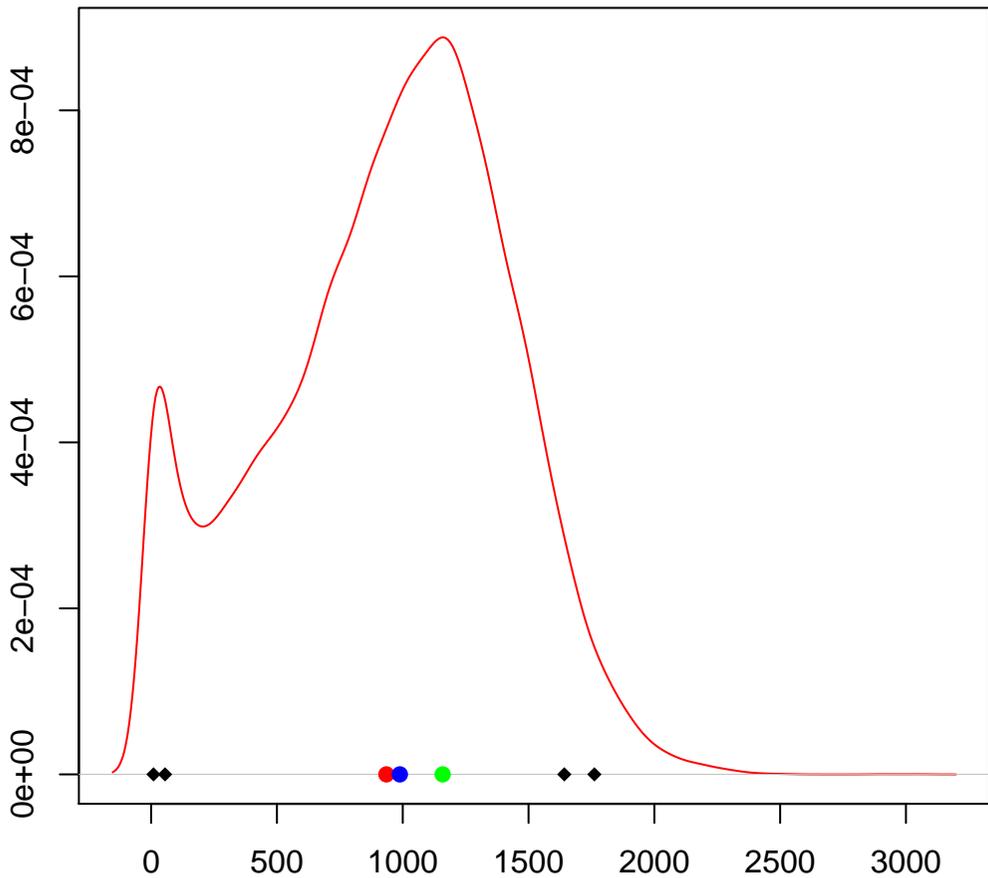
# VarA+VarD



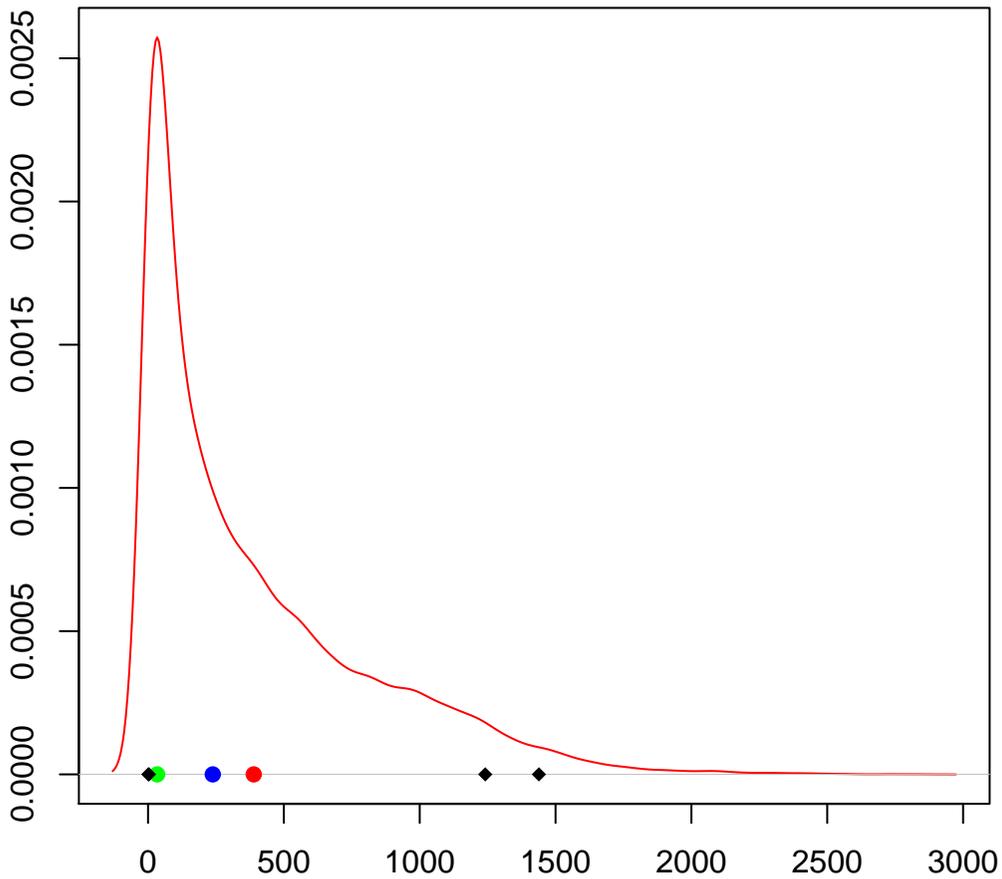
# VarA+VarD+VarC



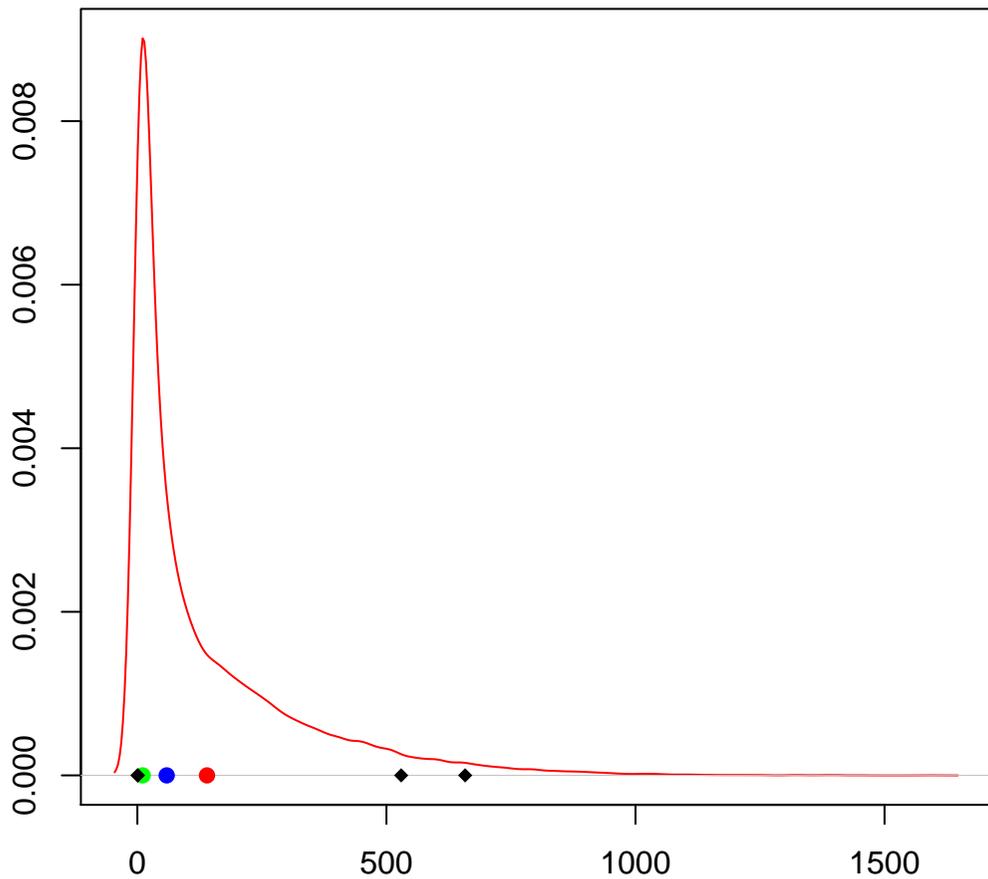
# VarA



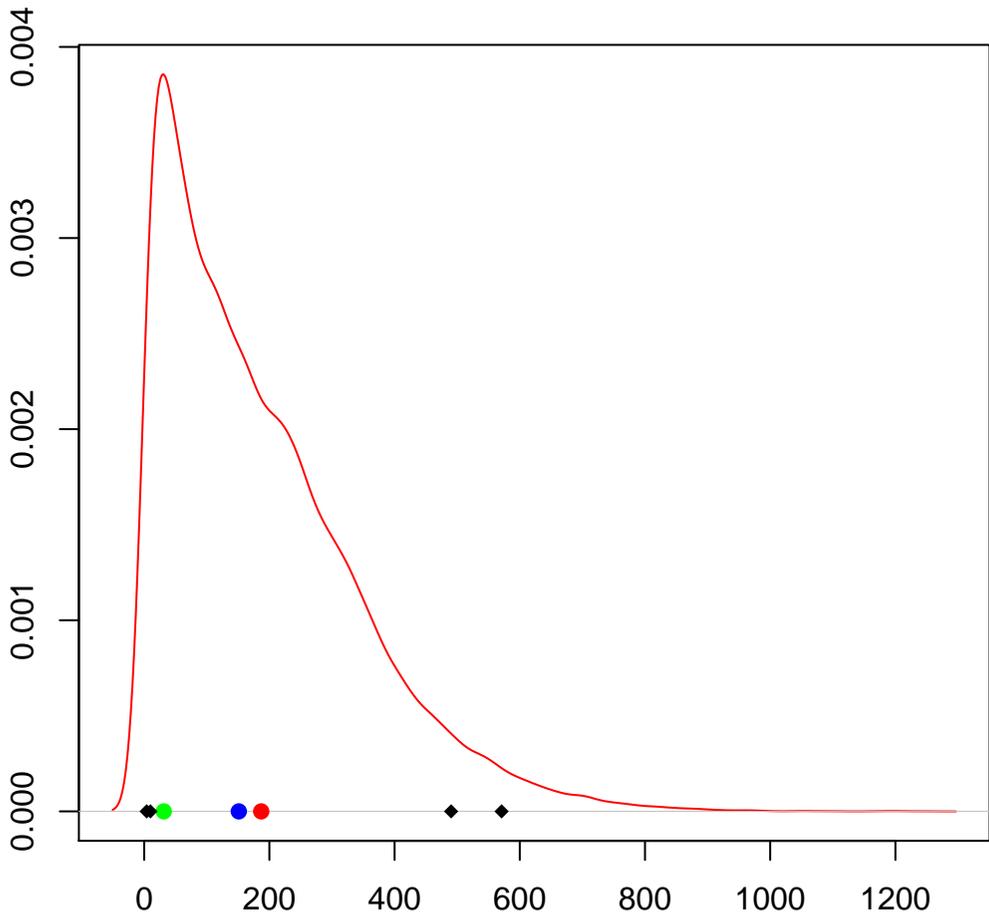
# VarD



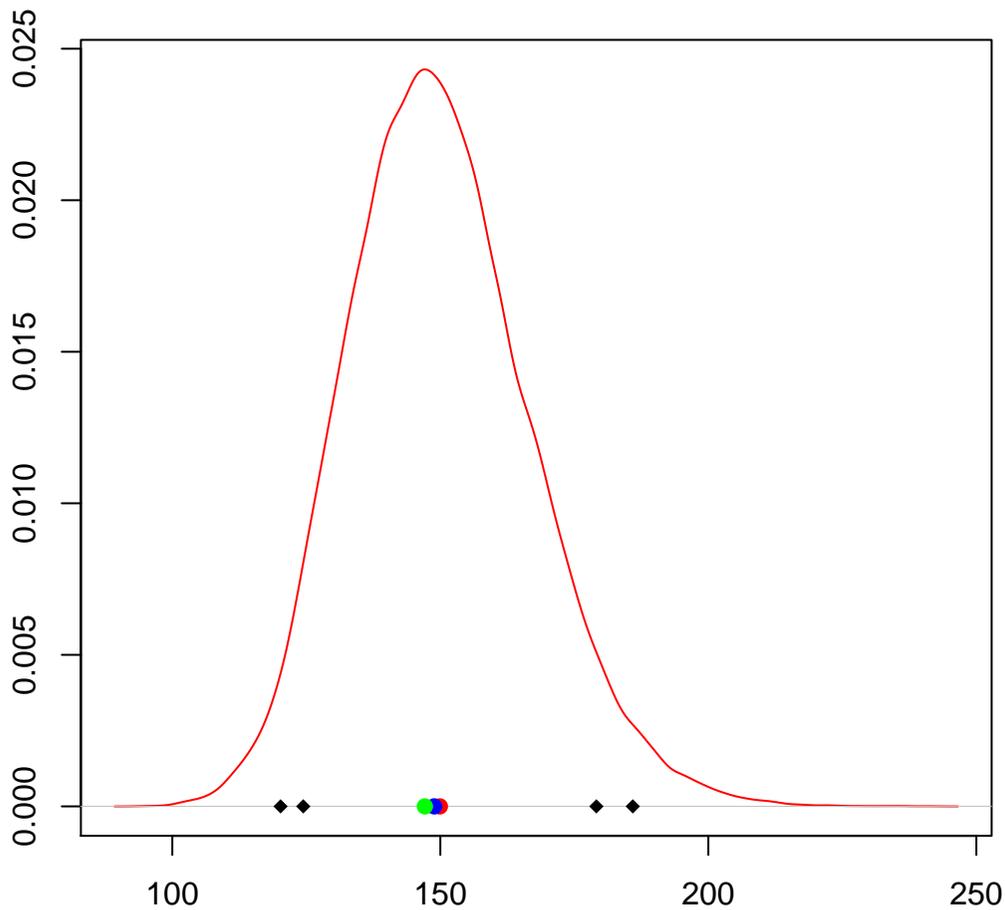
# VarC



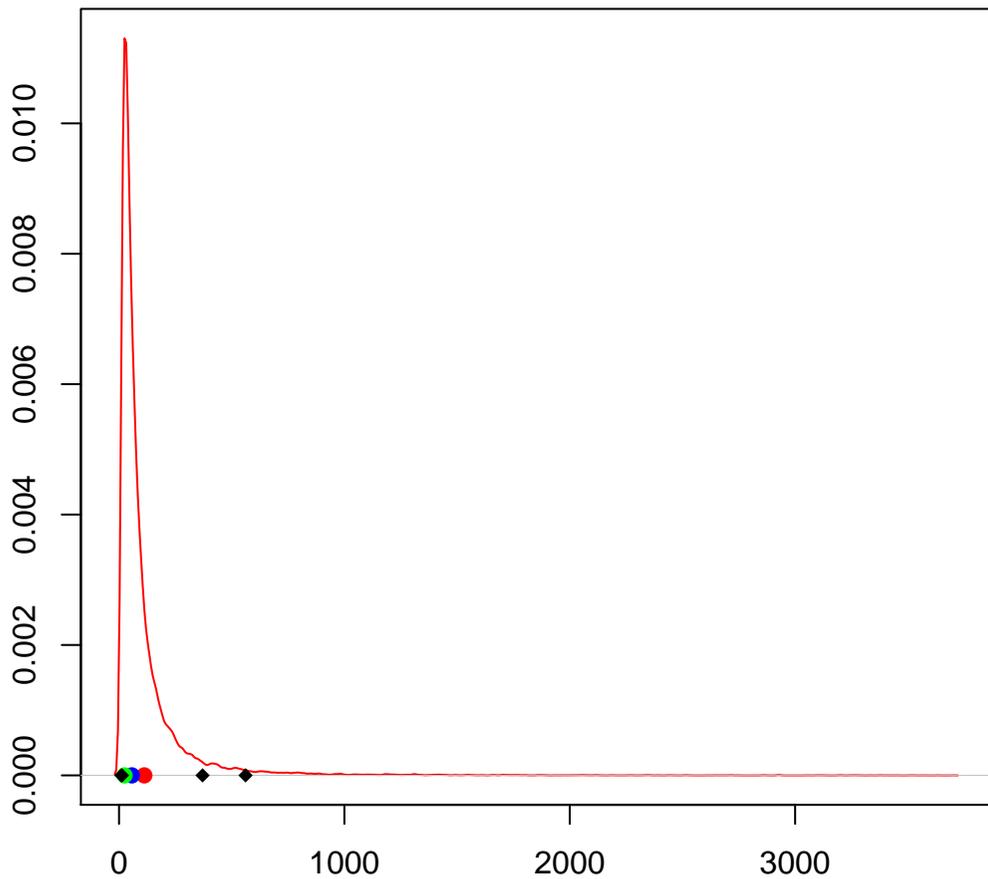
# VarE



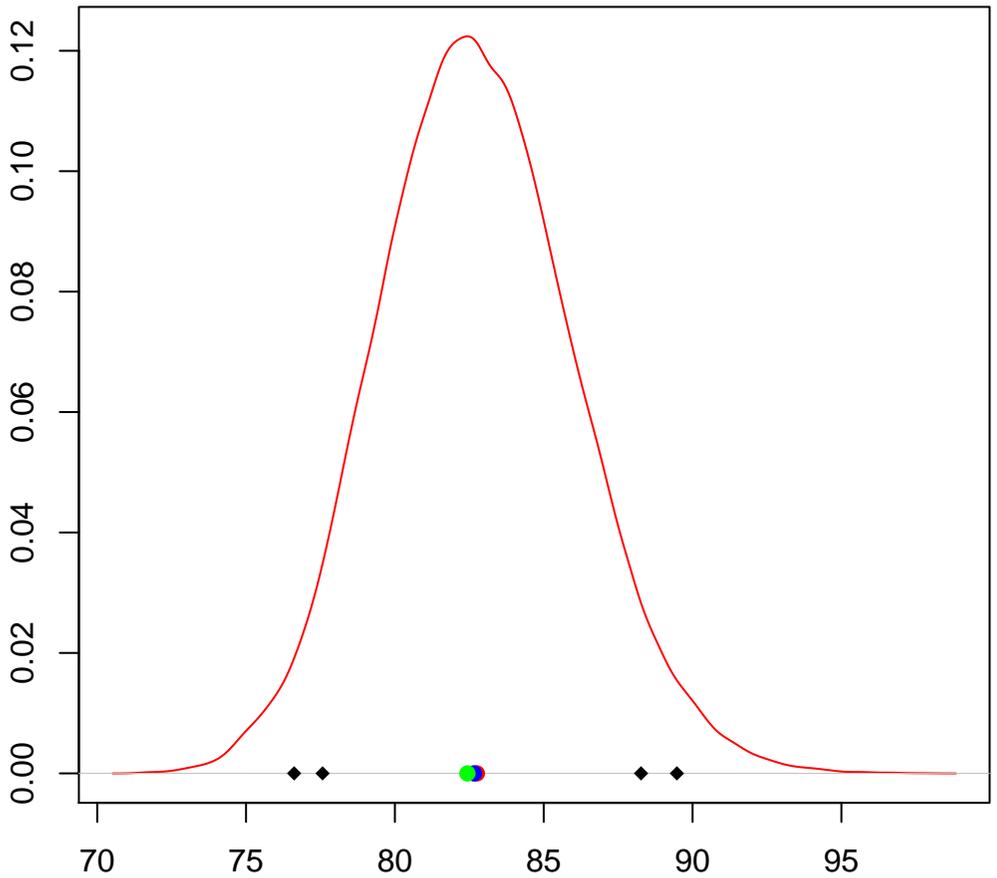
# VarAI



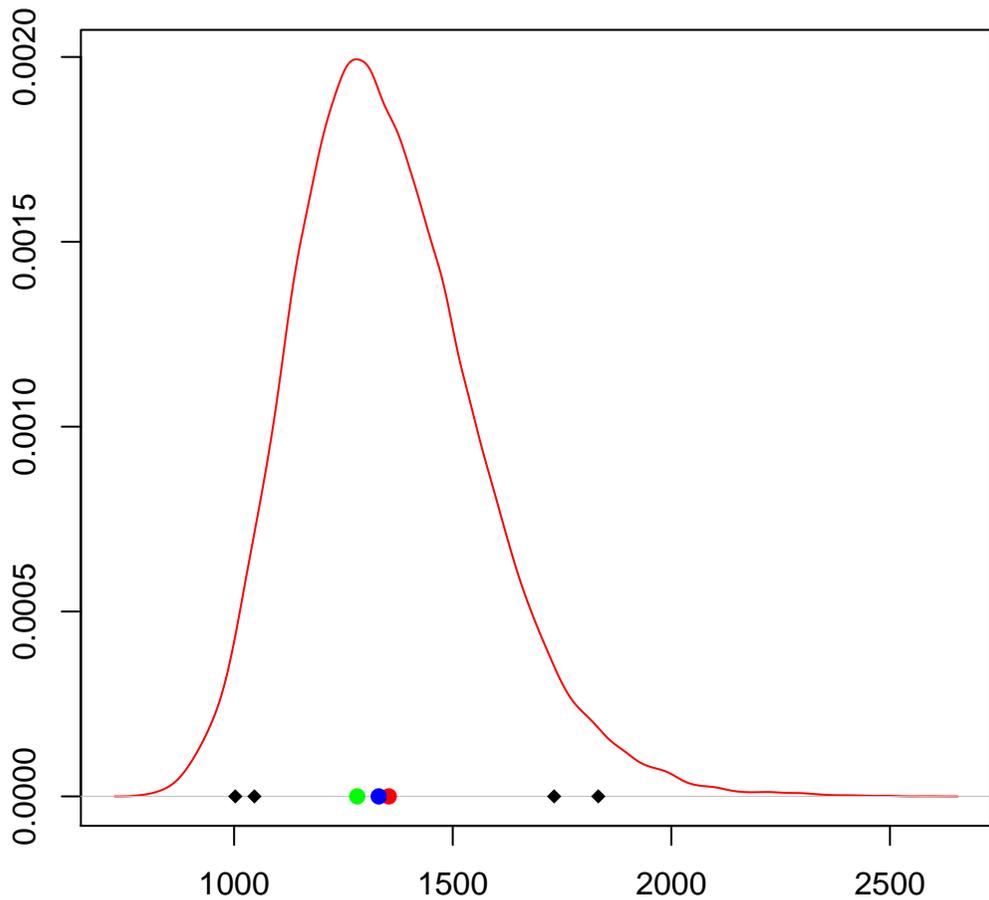
# VarB



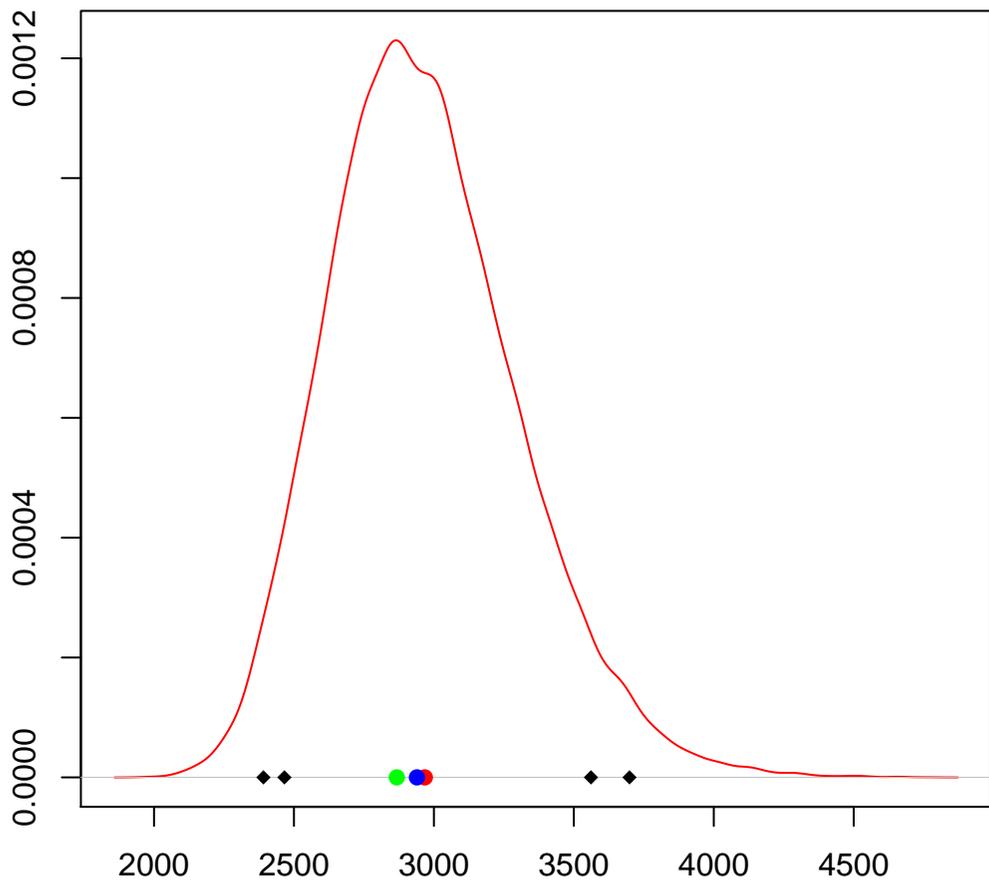
# VarEps



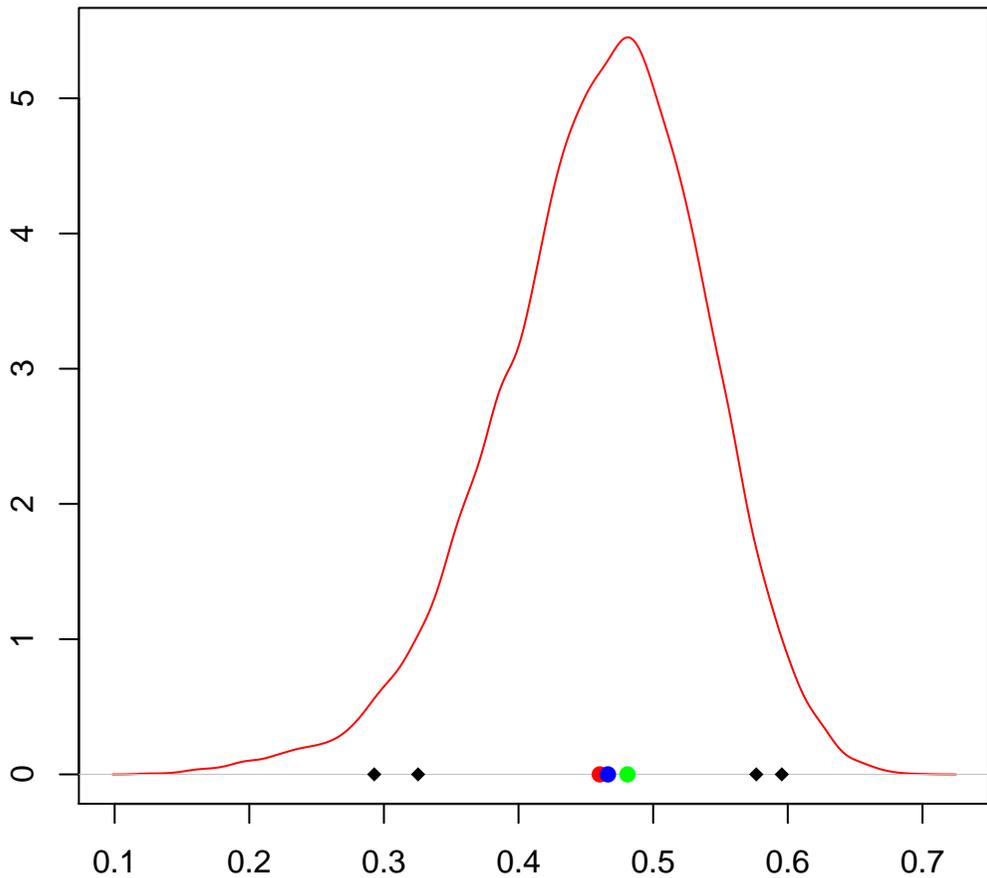
# VarE+VarV



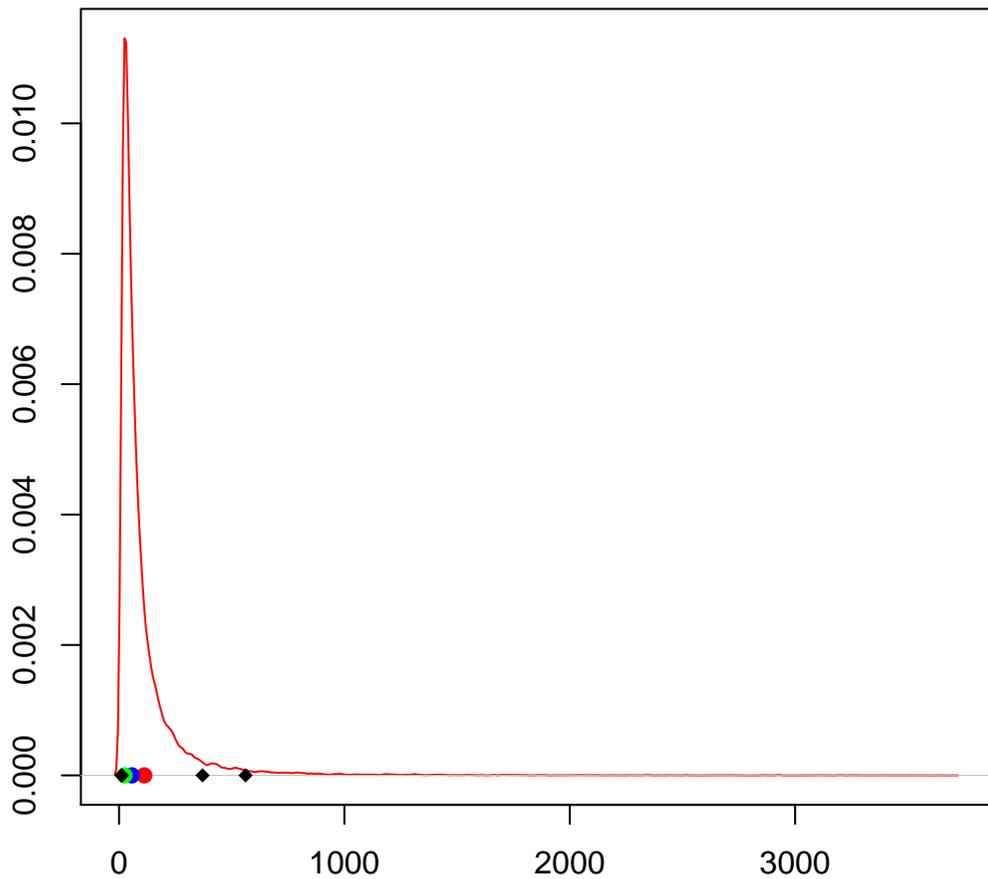
# VarA+VarD+VarC+VarE+VarV+VarAI



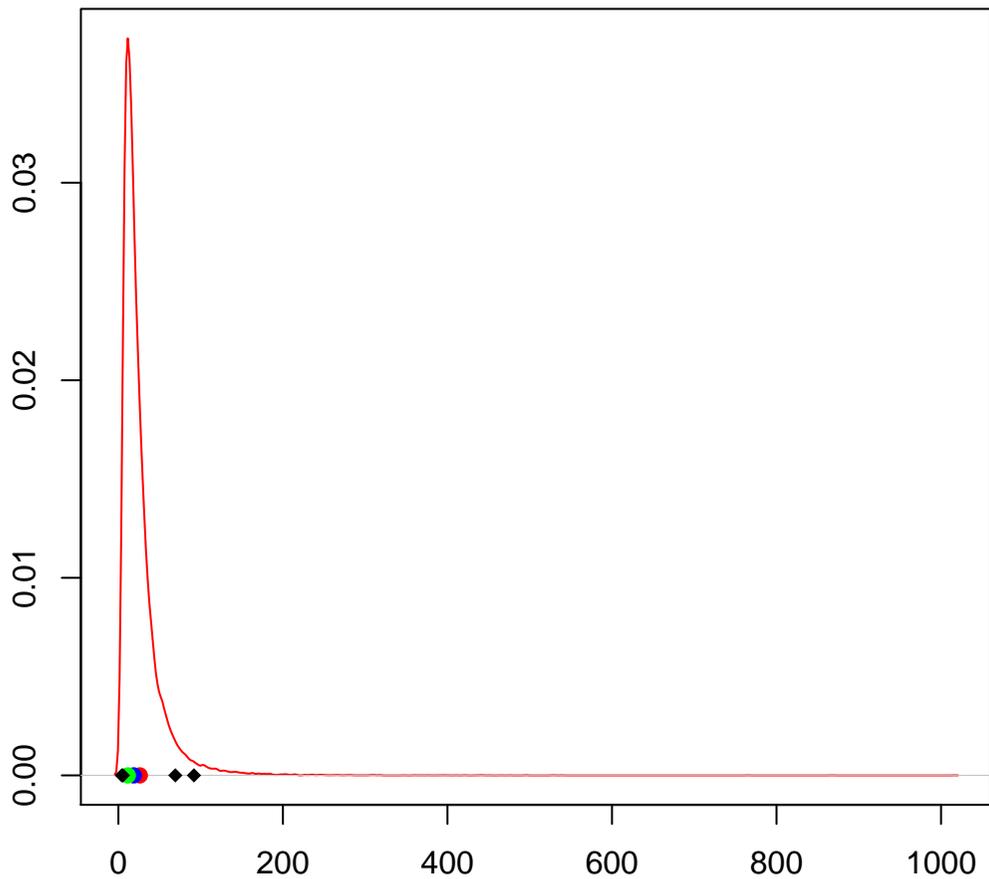
# Familiarity

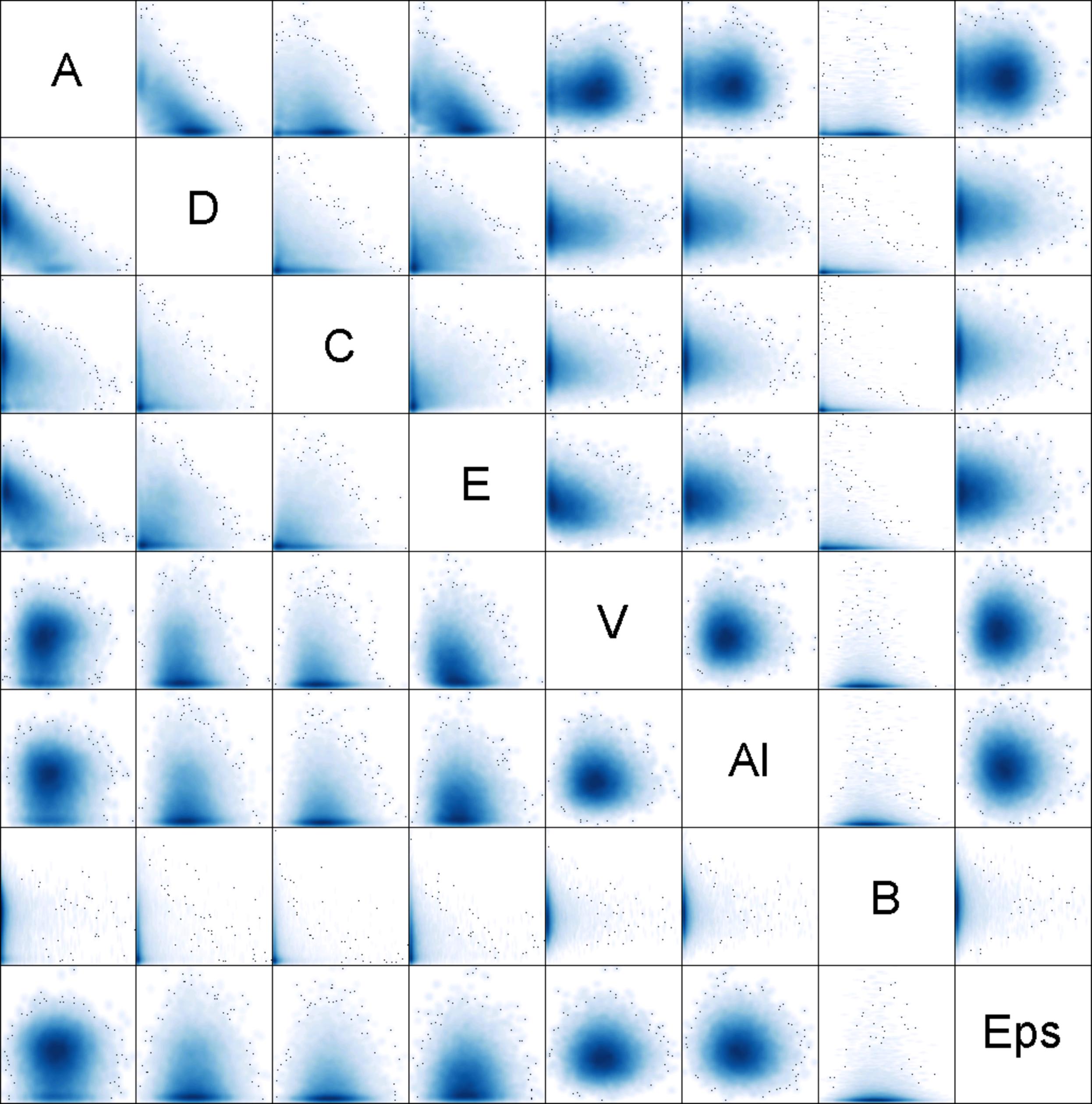


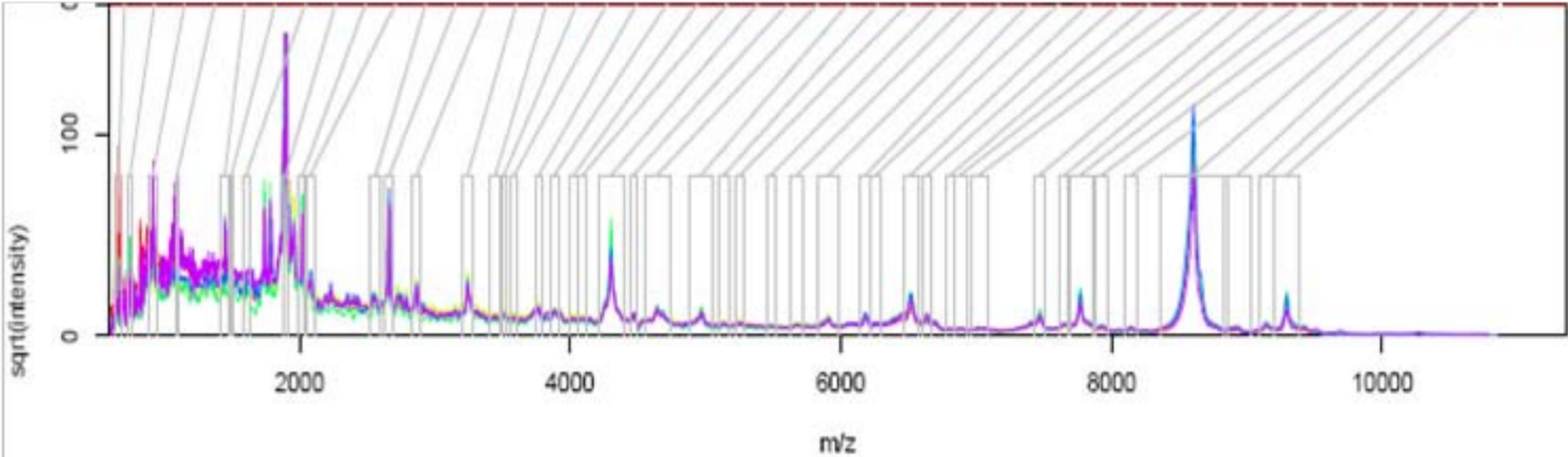
# VarB



# VarB





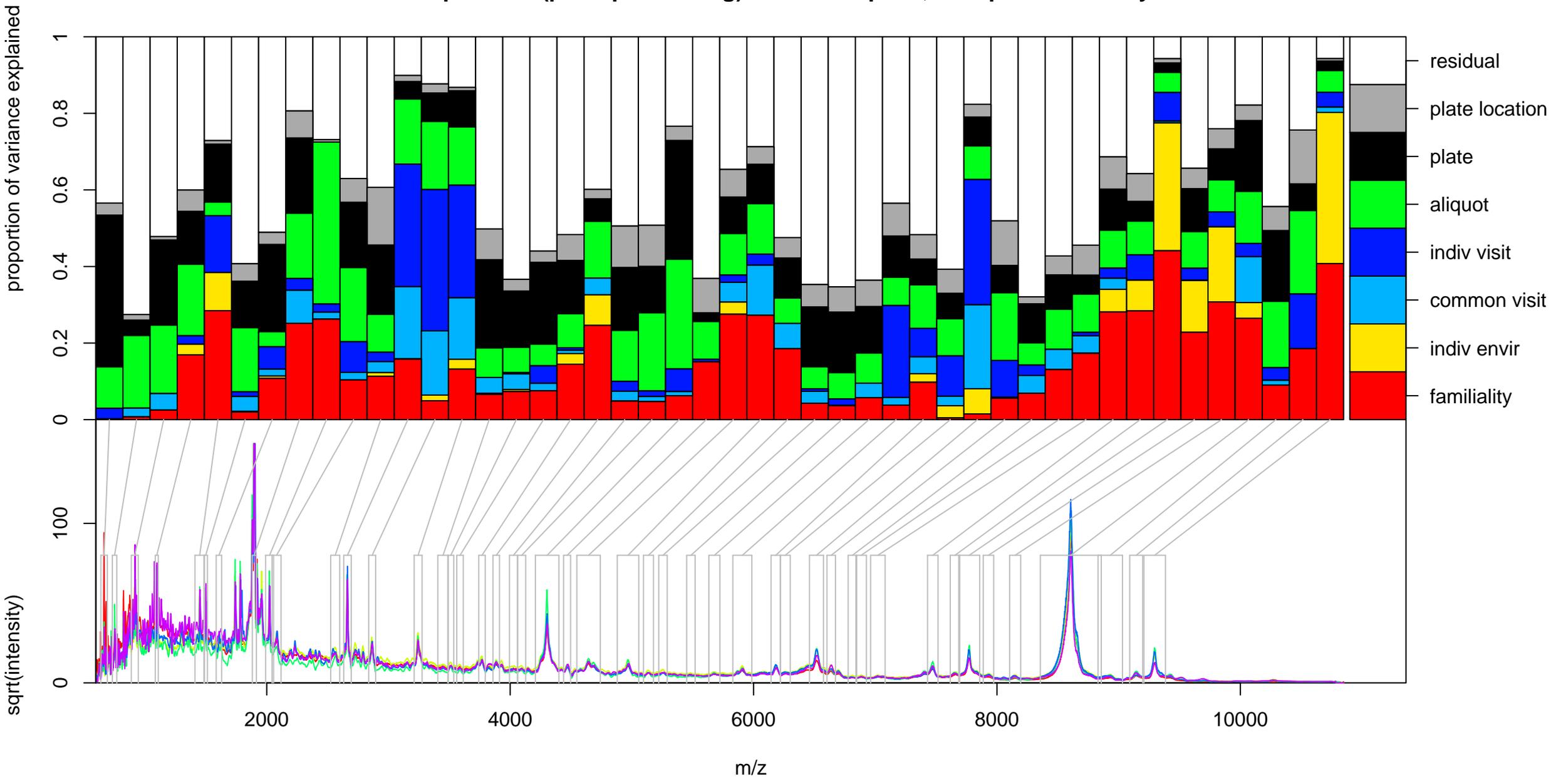


## Output

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- There are typically many peaks per spectra
- Our code does the spectral preprocessing, extracts peaks, runs the mcmc, and then reports posterior summary statistics

**imac bead type**  
**top: estimated variance components for each of 46 peaks summarised by sqrt(total)**  
**bottom: median spectrum (post-processing) from each plate, with peak summary intervals**



## Covariance Components models of association

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- We are interested in associating changes in molecular phenotype levels with changes in a clinical phenotype
- We have developed a new approach for this **when we have data on relateds**
- Consider a clinical phenotype,  $Z$  and molecular phenotype  $Y$
- A typical model would consider testing

$$\pi(Z|Y) \neq \pi(Z)$$

- However, it is interesting (we believe) to look for **genetic components of association**

- That is,

$$\pi(Z_{genetic}|Y_{genetic}) = \pi(Z_{genetic})$$

- We do this by investigating association between the genetical components of variation

- Consider the two phenotypes, one clinical and one molecular

$$Y_{ijkl} = \mu + a_{ij}^{(Y)} + d_{ij}^{(Y)} + c_i^{(Y)} + e_{ij}^{(Y)} + v_{ijk}^{(Y)} + l_{ijkl}^{(Y)} + b_{B(i,j,k,l)}^{(Y)} + \epsilon_{ijkl}^{(Y)}$$

$$Z_{ijkl} = \mu + a_{ij}^{(Z)} + d_{ij}^{(Z)} + c_i^{(Z)} + e_{ij}^{(Z)} + v_{ijk}^{(Z)} + l_{ijkl}^{(Z)} + b_{B(i,j,k,l)}^{(Z)} + \epsilon_{ijkl}^{(Z)}$$

- We can put a joint dependence structure on “interesting” components

- For example,

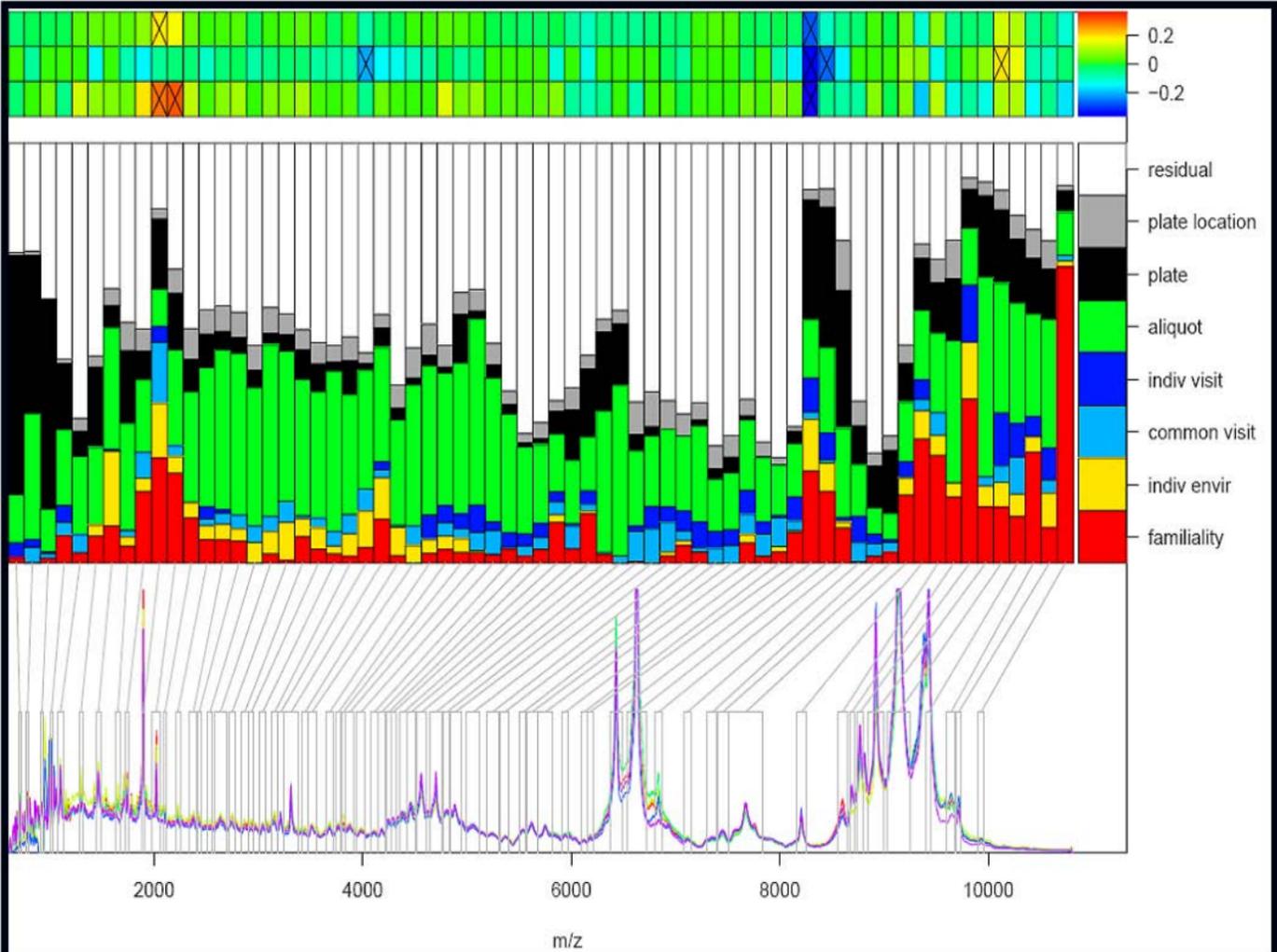
$$\{a^{(Y)}, a^{(Z)}\} \sim N(0, \rho_g \sigma_a^{(Y)} \sigma_a^{(Z)})$$

$$\{d^{(Y)}, d^{(Z)}\} \sim N(0, \rho_g \sigma_d^{(Y)} \sigma_d^{(Z)})$$

- with prior say

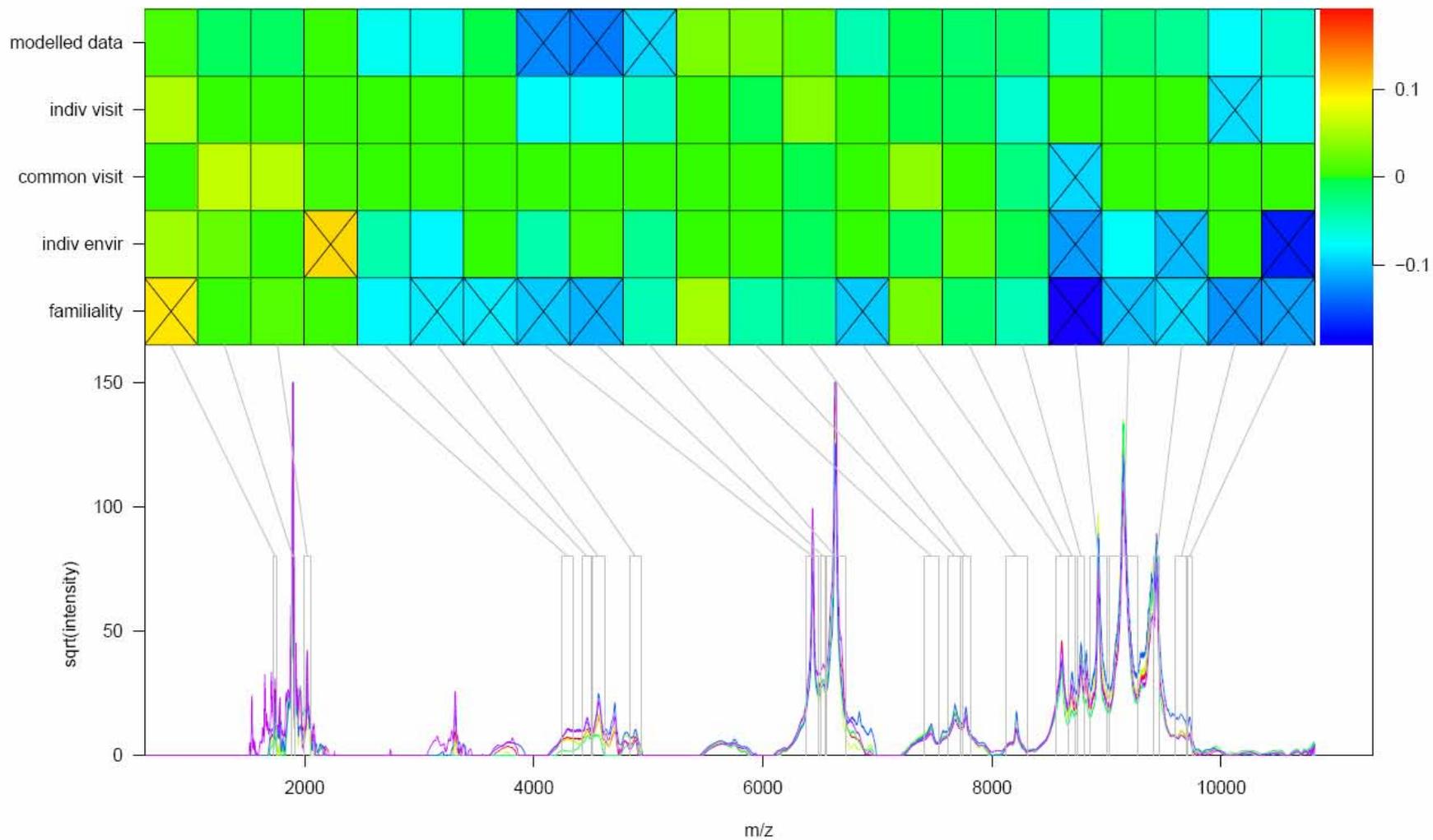
$$\pi(\rho_g) \sim U(-1, 1)$$

- and then investigate  $\pi(\rho_g | Y)$
- This looks for association in the **genetical axis of variation** between  $Z$  and  $Y$
- That is, in genetical projections orthogonal to that variability spanned by environmental and technical effects
- Summarise posterior mean associations



### c8 bead type

top: correlation of log(bmi) with each fitted random effect for each of 22 peaks summarised by sqrt(total); significantly non-zero correlation estimates marked with X; fdr = 0.05  
bottom: median spectrum (post-processing) from each plate, with peak summary intervals



## Summary

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- Information of relatedness allows us to separate out genetical from environmental factors in molecular phenotypes
- Bayesian framework very useful for what we do
- **Covariance components models** allow us to explore interesting axes of association
  - interested in extensions in graphical models/networks

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