Analyzing Cross-Plattform Consistency Using Tests Against Ordered Alternatives CAMDA Emerald Competition

#### Florian Klinglmueller<sup>1</sup> Thomas Tuechler<sup>2</sup>

<sup>1</sup>Core Unit for Medical Statistics and Informatics Medical University of Vienna florian.klinglmueller@meduniwien.ac.at <sup>2</sup>WWTF Chair for Bioinformatics BOKU University thomas.tuechler@boku.ac.at

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#### Introduction

#### Material and Methods

Experimental Design Methods

#### Exploratory Data Analysis

Total-RNA to Messenger-RNA Saturation

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#### Results

Monotone Genes Across Platform Normalization Effect

Discussion - Outlook Summary and Discussion Titration



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# Experimental Design:

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## Main Questions

Do the measured intensities reflect the titration?

- Agreement across platforms.
- Influence of normalization.

## Tests Against Order-Restricted Alternatives

- Dose-response studies
- 70's and 80's literature:
  - Barlow [1]
  - Robertson et al. [3]
- Microarray Application: Lin et al. [2]
- 5 Statistics: Marcus, Wilson, E2, M, ModifiedM

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• E2 most powerful  $\Rightarrow$  we use E2

## Test Null Hypothesis

We test the null hypothesis of equal means

$$H_{0,g}: \mu_{L,g} = \mu_{M1,g} = \mu_{M2,g} = \mu_{K,g}, \tag{1}$$

against the ordered alternatives

$$H_{1,g}^{up}: \quad \mu_{L,g} \le \mu_{M1,g} \le \mu_{M2,g} \le \mu_{K,g}, \tag{2}$$

$$H_{1,g}^{down}: \quad \mu_{L,g} \ge \mu_{M1,g} \ge \mu_{M2,g} \ge \mu_{K,g}, \tag{3}$$

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with at least one strict inequality.

▶ Main Principle: Isotonic Regression

Fitting Monotone Functions

### Isotonic Regression: Formulation

Isotonic Function  $\blacktriangleright$  Set  $\mathcal{T} := \{t_1, ..., t_n\}$  with order relation  $\blacktriangleright m(t_i)$  is called isotonic if  $t_i \leq t_j \Rightarrow m(t_i) \leq m(t_j)$   $\blacktriangleright \mathcal{F}(\mathcal{T})$ : all isotonic functions on  $\mathcal{T}$   $\blacktriangleright$  Direction has to be specified Isotonic Regression  $\blacktriangleright y_i = m(t_i) + \epsilon_i, m \in \mathcal{F}(\mathcal{T})$   $\blacktriangleright$  Least-squares fit:  $\hat{m} = \operatorname{argmin}_{m \in \mathcal{F}(\mathcal{T})} \sum_{i=1}^{n} (y_i - m(t_i))^2$ .

Example

$$\bullet \ \mathcal{T} = \{L \le M1 \le M2 \le K\}$$

$$\overline{y}_g(t_i) = m^{up}(t_i) + \epsilon_i$$

Some gene expressions:



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Upwards Trend

$$\blacktriangleright \ \mathcal{T} = \{L \le M1 \le M2 \le K\}$$

$$\overline{y}_g(t_i) = m^{up}(t_i) + \epsilon_i$$

Isotonic Regression for upwards trend:



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Downwards Trend

$$\mathcal{T} = \{L \ge M1 \ge M2 \ge K\}$$

$$\overline{y}_g(t_i) = m^{down}(t_i) + \epsilon_i$$

Isotonic Regression for downwards trend:



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## Statistic Definition of E2 Statistic

E2 (Barlow [1],Robertson et al. [3]):

$$\overline{E}_{01}^{2up} = 1 - \frac{\sum_{kj} (y_{kj} - \hat{m}^{up}(t_i))^2}{\sum_{kj} (y_{kj} - \overline{y})^2}, \qquad (4)$$

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Likelihood-ratio:

$$\overline{E}_{01}^{2up} = 1 - \frac{ESS}{TSS}$$

Capturing the Hierarchical Variance Structure

- Revisit the design hierarchy
- Now we add a new level: Normalization



# Normalizations

Baseline vs. Quantile Normalization

## Both widely used

## **Baseline Normalization**

Align per array medians

1. From each array remove array-wise median

2. To each array add overall median

Removes systematic location shifts

## Quantile Normalization

Align order statistics

- 1. Per array reduce expressions to ranks
- 2. Per array reassign ranks to quantiles from mean distribution (means of order statistics)

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Removes any systematic disturbance that keeps the order

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Capturing the Hierarchical Variance Structure

- Revisit the design hierarchy
- ▶ We want *p*



Inverse Normal Method

Combine one-sided *p*-values:

$$p_g^{C,up} = 1 - \Phi(\frac{1}{\sqrt{N}} \sum_i \Phi^{-1}(1 - p_{ig}^{up})),$$
 (5)

▶  $p_g^{C,down}$  analogue

- uniformly distritibuted conservative one-sided p-values
- Bonferroni correct directional decision:  $p_g^C = 2\min(p_g^{C,up}, p_g^{C,down}).$

Per Animal *p*-Values





Per Animal *p*-Values

- ▶ 6 Animals × 3 Platforms × 2 Normalizations → 36 times  $P_{Norm,Plat,ig}^{up}$ ,  $P_{Norm,Plat,ig}^{down}$ ,  $P_{Norm,Plat,ig}$
- Combine the 6 × 6 p<sup>up</sup><sub>Norm,Plat,ig</sub>, p<sup>down</sup><sub>Norm,Plat,ig</sub> to get get 6: p<sup>CPlat</sup><sub>Norm,g</sub>, p<sup>CPlat</sup><sub>Norm,g</sub>, and p<sup>CPlat</sup><sub>Norm,g</sub>

   Combine the 3 p<sup>CPlat</sup><sub>Norm,g</sub>, p<sup>CPlat</sup><sub>Norm,g</sub> to get 2: p<sup>CNorm,up</sup><sub>Norm,down</sub>



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Per Animal p-Values

 6 Animals × 3 Platforms × 2 Normalizations → 36 times <sup>up</sup><sub>Norm,Plat,ig</sub>, p<sup>down</sup><sub>Norm,Plat,ig</sub>, PNorm,Plat,ig</sub>
 Combine the 6 × 6 p<sup>up</sup><sub>Norm,Plat,ig</sub>, p<sup>down</sup><sub>Norm,Plat,ig</sub> to get get 6: <sup>CPlat</sup>,up, p<sup>CPlat</sup>,down, and p<sup>CPlat</sup><sub>Norm,g</sub>
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- Comptute one sided permutation test *p*-values for each animal, on each platform seperately with Quantile - and Baseline - normalized data.
- Combine per animal tests from each plaform.
- Combine per platform tests from each normalization.

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# Finally!

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# Exploratory Analysis

Distribution of Group Means on Raw Data



- Location-shift
- Higher messenger-RNA content in kidney?
- Both normalization methods remove any visible trends in location

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- Baseline
- ► Quantile also in scale

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- Baseline
- Quantile also in scale

Relationship between Increases



## Relationship between first/second increase

 Scatterplot - Illumina: Trends not linear; When first increase large then last increase small and vice versa

- Scatterplot Agilent
- Scatterplot -Affymetrix
- Rightmost point

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- Lowest point
- Saturation?

Relationship between Increases

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## Illumina

- Relationship between first/second increase
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Relationship between Increases

# 0 ZM-3 M-M2 -6 5 10 0 M1-L

Agilent

- Relationship between first/second increase
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Relationship between Increases

# $\begin{array}{c} 0 \\ -1 \\ -1 \\ -2 \\ -3 \\ -4 \\ -5 \\ -6 \\ -6 \\ 0 \\ 2 \\ 4 \\ 6 \\ M1-L \end{array}$

## Affymetrix

- Relationship between first/second increase
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Relationship between Increases



#### NM\_052802

- Relationship between first/second increase
- Scatterplot Illumina: Trends not linear; When first increase large then last increase small and vice versa
- Scatterplot Agilent
- Scatterplot -Affymetrix
- Rightmost point
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# $\begin{array}{c} 0 \\ -1 \\ -1 \\ -3 \\ -3 \\ -4 \\ -5 \\ -6 \\ -6 \\ 0 \\ 2 \\ 4 \\ 6 \\ M1-L \end{array}$

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Relationship between Increases



#### NM\_022519

- Relationship between first/second increase
- Scatterplot Illumina: Trends not linear; When first increase large then last increase small and vice versa
- Scatterplot Agilent
- Scatterplot -Affymetrix
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## Affymetrix

# Test Setup

## Settings

- R package IsoGene provided by Lin et al.
- 20000 permutations (1 week on Cluster)
- 2 Normalization Methods × 3 Platforms × 6 Animals

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- 6111 well annotated genes available on all platforms
- remove one animal from Illumina data
- ► Family Wise Error: Bonferoni-Holm

# Proportions of Significant Genes

General Overview



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► Quantile

# Proportions of Significant Genes

General Overview



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# Agreement Between Platforms

Number of Genes



 Fleiss' κ-coefficient - agreement across platforms using FWR adjusted combined *p*-Vaues

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- Quantile Normalisation: .52
- Baseline Normalisation: .37

## Agreement Between Normalizations

Number of Genes significant



Fleiss  $\kappa$ -coefficient: .57

- around 2 times more significant genes exclusive to baseline than to quantile normalized data
- more than 97% of genes exclusive to baseline normalized data are upregulated
- up-down in quantile exclusive genes 40:60

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## Data

## Substantial number of genes show significant monotonicity

- Across platform agreement exceeds chance levels
- Agreement on baseline normalized data is worse
- Baseline noramlized data shows more upward trends incomplete removal of total/messenger-RNA effect
- Genes exclusively significant in baseline data are mostly upward trends

## Methods

- Isotonic regression as a means to detect monotonic trends
- *p*-Value combination as a means to compare results from differnt platforms.

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## Thanks

- MSI Martin Posch
- Statistic Univie: Cluster

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## References

- [1] Richard E. Barlow. *Statistical Inference Under Order Restrictions.* John Wiley and Sons Ltd, 1972.
- [2] D. Lin, Z. Shkedy, D. Yekutieli, T Burzykowski, H. Gaehlmann, A. Bondt, T. Perera, T. Geerts, and L. Bijnens. Testing for trends in dose-response microarray experiments: a comparison of several testing procedures, multiplicity and resampling-based inference. *Statistical Applications in Genetics and Molecular Biology*, 2007.
- [3] Tim Robertson, F. T. Wright, and R. L. Dykstra. Order Restricted Statistical Inference. John Wiley & Sons Inc, 1988.

# Thank you for your attention